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Chemical Constituents from the roots of Ranunculus ternatus

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Two new glycosides named as ternatoside A (1) and ternatoside B (2), with four known constituents sternbin (3), methylparaben (4), 4-O-D-glucopyranosyl-p-coumaric acid (5) and linocaffein (6) were isolated from the roots of *Ranunculus ternatus*, The structures of new compounds were determined by 1D and 2D NMR, MS techniques, and chemical methods.

Keywords: Ranunculus ternatus; Ternatoside A; Ternatoside B

1. Introduction

Ranunculus ternatus Thunb. is a plant of genus *Ranunculus* used for the treatment of tuberculosis [1]. Some fatty acid esters and γ -keto- δ -valerolactone have been isolated from this plant [2,3]. In this paper, we reported the isolation and structural elucidation of two new glycosides, ternatoside A (1) and B (2), with other four known constituents sternbin (3), methylparaben (4), 4-O-D-glucopyranosyl-p-coumaric acid (5) and linocaffein (6). The structures were identified on the basis of spectroscopic and chemical methods.

2. Results and discution

Compound 1 was obtained as a colorless gum, and gave positive result to Molish test. In the positive and negative ESIMS, it showed quasi-molecular ion peaks at m/z 373.2 [M + Na]⁺, 189.2 [M - 162 + H]⁺ and 349.3 [M - H]⁻, respectively. The molecular formula $C_{15}H_{26}O_9$ was determined by HRFABMS and ¹³CNMR. Glucose was detected after the acid hydrolysis and compared with authentic sample on TLC. The ¹H, ¹³CNMR and HMQC spectra indicated that this compound possesses three methylenes, one O-butyl, one carbonyl of ketone, one carbonyl of ester, and one glucosyl group (table 1). The structure of 1 was determined on the basis of ¹H-¹HCOSY and HMBC spectra. In HMBC spectrum of 1,

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Table 1. NMR spectral data of 1 (¹H, 500 MHz; ¹³C, 125 MHz; δ ppm, J Hz, in DMSO-*d*₆).

No.	δ_H	δ_C	No.	δ_H	δ_C
1		172.9	9	0.87(t, 3H, 7.5)	14.0
2	2.48(t, 2H, 6.5)	27.7	1'	4.91(d, 1H, 7.5)	103.3
3	2.78(t, 2H, 6.5)	34.1	2'	3.04(t, 1H, 7.5)	74.0
4		207.9	3'	3.17(t, 1H, 7.5)	77.2
5	4.30, 4.22(d, 2H, 17.0)	73.7	4′	3.07(t, 1H, 7.5)	70.7
6	3.99(t, 2H, 7.5)	64.2	5'	3.25(m, 1H, 7.0)	77.5
7	1.52(m, 2H, 7.5)	30.8	6'	3.99, 3.66(dd, 2H, 13.5, 7.0)	61.7
8	1.31(m, 2H, 7.5)	19.1			

¹³C⁻¹H long-range correlations were found between H-2, H-3, H-6 and C-1; H-2, H-3, H-5 and C-4; H-5 and C-3; The anomeric proton of glucosyl group at δ 4. 91 (d, 1H, *J* = 7.5 Hz) was correlated to C-5 of aglycone (figure 1). In ¹H⁻¹HCOSY spectrum of **1**, correlations were found between H-2 and H-3; H-6 and H-7; H-7 and H-8; H-8 and H-9. The anomeric configuration of the glucose was deduced to be β-anomer from the coupling constant of the anomeric proton. Thus compound **1** was elucidated as 4-carbonyl-(O-β-D-glucopyranosyl)pentanoic acid-1-O-butyl ester, named as ternatoside A.

Compound 2 was obtained as a brown gum, and gave positive result to Molish test. In the positive and negative ESIMS, it showed quasi-molecular ion peaks at m/z 439.2 [M + Na]⁺, $255.2 [M - 162 + 1]^+$ and $415.3 [M - H]^-$, respectively. Its molecular formula $C_{19}H_{28}O_{10}$ was established by HRFABMS and ¹³CNMR. Glucose was detected after the acid hydrolysis and compared with authentic sample on TLC. The ¹H, ¹³CNMR and HMOC spectra indicated that this compound possesses 19 carbon signals including one methylene, one methine, one phenyl, one O-butyl, one glucosyl group, one carbonyl of ester groups (table 2). In the HMBC of compound 2, ${}^{13}C-{}^{1}H$ long-range correlations were found between H-2, H-3, H-4 and C-1; H-2, H-2', H-6' and C-3; H-2, H-3, H-2', H-5', H-6' and C-1'; H-2', H-5', H-6' and C-4'; The anomeric proton of glucosyl group at δ 4.61(d, 1H, J = 7.5 Hz) was correlated to C-4' of phenyl (figure 1). In ${}^{1}H^{-1}HCOSY$ spectrum of 2, correlations were found between H-2 and H-3; H-5' and H-6'; H-4 and H-5; H-5 and H-6; H-6 and H-7. The anomeric configuration of the glucose was deduced to be β -anomer from the coupling constant of the anomeric proton. A comparison of the ¹³CNMR spectra of 2 with those of the known (R)-3- $[3-hydroxy-4-(-O-\beta-D-glucopyranosyl)-phenyl]-2-hydroxypropanoic acid [4], showed very$ similar δ value, except for C-1 of **2** up-shifted for 10.90 ppm. Combining with the aid of HMBC correlation, it suggested that the carboxy of **2** have been esterified by butanol. Optical rotation of 2 was also compared with the known (R)-3-[3-hydroxy-4-(-O- β -Dglucopyranosyl)-phenyl]-2-hydroxypropanoic acid, indicating compound 2 has a (R)-form. Therefore, compound 2 was elucidated as (R)-3-[3-hydroxy-4-(-O- β -D-glucopyranosyl)phenyl]-2-hydroxypropanoic acid butyl ester, named as ternatoside B.



Figure 1. Structures and Key HMBC correlations of 1 and 2.

No.	δ_H	δ_C	No.	δ_H	δ_C
1		174.3	4′		146.0
2	4.16 (t, 1H, 6.0)	72.0	5′	6.67(d, 1H, 6.5)	116.1
3	2.81,2.69(dd, 2H, 14.5,6.0)	40.0	6′	6.69(dd, 1H, 6.5, 2.5)	124.4
4	3.96(t, 2H, 7.0)	64.3	1″	4.61(d, 1H, 7.5)	103.2
5	1.49(m, 2H, 7.0)	30.8	2″	3.27(t, 1H, 7.5)	74.0
6	1.26(m, 2H, 7.0)	19.2	3″	3.25(t, 1H, 7.5)	76.6
7	0.89(t, 3H, 7.0)	14.2	4″	3.18(t, 1H, 7.5)	70.3
1'		129.0	5″	3.29(m, 1H, 6.5)	77.8
2'	6.95(d, 1H, 2.5)	118.7	6″	3.72,3.63(dd, 2H, 11.7,6.5)	61.4
3′	··· · /	145.5			

Table 2. NMR spectral data of 2 (¹H, 500 MHz; ¹³C, 125 MHz; δppm , J Hz, in DMSO- d_6).

3. Experimental

3.1 General experiment procedures

Melting points were measured on a Fisher-Johns apparatus and are uncorrected. Optical rotations were obtained on a Perkin-Elmer 341 polarimeter. UV spectra were measured by a Waters-2996 detector. IR spectra were recorded on a Perkin-Elmer 983G spectrometer. NMR spectra were measured on a Bruker AM-500 (500 MHz) instrument. FABMS were obtained on a Zabspec E spectrometer; ESIMS were obtained on an Esquire-LC00054 spectrometer. For column chromatography, silica gel (200–300 mesh, Qingdao Haiyang Chemical Co.) was used. TLC and HPTLC (silica gel GF₂₅₄ precoated plates, Qingdao Haiyang Chemical Co.) detection was obtained by spraying 10% H_2SO_4 followed by heating.

3.2 Plant material

The *Ranunculus ternatus* was collected from Henan province of China in October 2002, and identified by Dr. Xue-Sen Wen, School of Pharmaceutical Sciences, Shandong University. A voucher specimen is deposited in Department of Chinese Medicine Sciences & Engineering, Zhejiang University.

3.3 Extraction and isolation

The dried powdered plant materials (10 kg) were refluxed with 70% EtOH twice, after removal of the solvent by evaporation, the combined extracts were partitioned between H₂O and petroleum ether, CHCl₃, EtOAc and *n*-BuOH, successively. The EtOAc extract (170 g) was chromatographed on Si gel column, eluting with CHCl₃–MeOH (MeOH contain 5% H₂O) from 100 to 50:50 in a gradient manner divided into 15 fractions. Fraction 3(CHCl₃– MeOH 95:5) (0.7 g) was separated on Si gel column, using petroleum ether-EtOAc (87:13) as eluent to afford **3**(26 mg), fraction 5(CHCl₃-MeOH 90:10) (2.3 g) was separated on Si gel column, using petroleum ether–EtOAc (82:18) as eluent to afford **4**(280 mg), Fraction 8(CHCl₃–MeOH 80:20) (13.0 g) was separated on Si gel column, using CHCl₃–MeOH (MeOH contain 5% H₂O) (86:14) as eluent to afford **5** (19 mg) and **6**(35 mg), Fraction 11(CHCl₃/MeOH 70:30) (7.7 g) was separated on Si gel column, using CHCl₃–MeOH (MeOH contain 5% H₂O) (77:23) as eluent to afford **1** (22 mg) and **2** (16 mg). J-K. Tian et al.

3.3.1 Compound 1. Colorless gum (MeOH), ¹H NMR (DMSO-*d*₆, 500 MHz) and ¹³C NMR (DMSO-*d*₆, 125 MHz) see table 1; positive ESIMS *m*/*z* 373.2 $[M + Na]^+$, 189.2 $[M - 162 + H]^+$; negtive ESIMS *m*/*z* 349.3 $[M - H]^-$; HRFABMS *m*/*z* 373.1426 $[M + Na]^+$ (calcd for C₁₅H₂₆O₉Na, 373.1475).

3.3.2 Compound 2. Brown gum (MeOH), $[\alpha]_D^{20} - 41.20$ (c 0.50, MeOH); ¹H NMR (DMSOd₆, 500 MHz) and ¹³C NMR (DMSO-d₆, 125 MHz) see table 2; positive ESIMS *m*/*z* 439.2 [M + Na]⁺, 255.2 [M - 162 + 1]⁺; negtive ESIMS *m*/*z* 415.3 [M - H]⁻; HRFABMS *m*/*z* 439.1551 [M + Na]⁺ (calcd for C₁₉H₂₈O₁₀Na, 439.1580).

3.3.3 Compound 3. Yellow needle crystal (CH₃Cl), mp $221-223^{\circ}$ C, positive ESIMS *m/z* 325.3 [M + Na]⁺, 303.3 [M + H]⁺. By comparison of NMR and UV data with those of the literature value [5], it was identified as sternbin (**3**).

3.3.4 Compound 4. White needle crystal (CH₃Cl), mp 129–131°C, IR (KBr) ν_{max} cm⁻¹ 3500, 3200, 2960, 2870, 1740, 1700, 1680, 1480, 1320 cm⁻¹; ¹H NMR (DMSO-*d*₆, 500 MHz) δ 9.62 (1H, s, COOH), 7.22 (2H, d, *J* = 6.3 Hz, H-2, 6), 6.57 (2H, d, *J* = 6.3 Hz, H-3, 5), 4.64 (3H, s, Me); ¹³C NMR (DMSO-*d*₆, 125 MHz) δ 177.7(COO), 157.2(C-1), 152.8(C-4), 122.0(C-2, 6), 111.9 (C-3, 5), 64.6(Me); positive ESIMS *m*/*z* 175.2 [M + Na]⁺, 153.1 [M + H]⁺. The above data revealed compound **4** was methylparaben.

3.3.5 Compound 5. Brown gum (MeOH), IR (KBr) ν_{max} cm⁻¹ 3400, 3300, 2850, 1770, 1690, 1670, 1320, 1200, 1030 cm⁻¹; positive ESIMS *m*/*z* 349.3 [M + Na]⁺, 327.4[M + H]⁺. By comparison of NMR data with those of the literature value [6], it was identified as 4-O-D-glucopyranosyl-p-coumaric acid (**5**).

3.3.6 Compound 6. Brown gum (MeOH), IR (KBr) ν_{max} cm⁻¹ 3450, 3280, 2860, 1770, 1690, 1670, 1330, 1210, 1020 cm⁻¹; positive ESIMS m/z 355.3 [M + Na]⁺, 343.3 [M + H]⁺. By comparison of NMR data with the literature value [7], it was identified as linocaffein (**6**).

3.3.7 Acid hydrolysis of 1–2 and 5–6. Compounds of 1–2 and 5–6 (each 5 mg) were refluxed with 5%HCl in MeOH (10 mL) for 5 h, each mixture was diluted with H₂O, neutralized with Na₂CO₃. The neutral hydrolysate revealed the presence of glucose by HPTLC [CH₃Cl–MeOH–H₂O (65:35:10) lower layer] when compared with authentic sample (Sigma).

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